



Enhanced gene therapy with TRAIL by bispecific antibody immuno therapy in advanced prostate and pancreatic cancer

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Introduction

Patients with advanced pancreatic or prostate cancer have a poor survival rate and new therapeutic strategies are needed. The overall aim of the project is the establishment and functional analysis of a new approach for the treatment of advanced prostate and pancreatic cancer. This approach will utilize a combination of the death ligand TRAIL as a pro-apoptotic agent over-expressed by T-cells and bispecific antibodies (bsAbs) targeting EpCAM on tumor cells and CD3 T-cells. The novel aspect is the combination of genetic modification with immune therapy. The lentiviral vectors allow the modification of lymphocytes to over-express the death ligand TRAIL (TNF-related apoptosis

inducing ligand). The bispecific antibodies (bsAbs) binds with one arm the T cell via CD3 and with the second arm the tumor surface antigen EpCAM, thereby bringing tumor and effector cell in closer contact. We analyzed these features in established primary prostate (PC-3) and pancreatic (BxPC-3) carcinoma cell lines, in in vitro three-dimensional tumor reconstructs and in mouse xenografts.



PBMCs were isolated from fresh healthy donor's blood, and following activation with IL-2 and OKT3 were transduced with lentiviral TRAIL. The activation with IL-2 and UK13 were transduced with lentiviral TRAIL. The chosen vector (pV3P2AE) is a lentiviral SIN vector of the 2nd generation, expressing TRAIL and EGFP under the control of a tet-activator. Transduction was done in 2 cycles on day 1 and 2 with MO110 each. Efficiency and TRAIL overexpression was verified 3 days post transduction via FACS analysis. All cells were used at this day in evanements.

Apoptosis induction in 3D tumor reconstructs





To evaluate the efficacy of bispecific EpCAMxCD3 antibodies, we established an *in vitro* 3D tumour reconstruct system. To mimic tumour microenvironment, a mix of transduced lymphocytes were co-cultu together with tumour cells and fibroblasts in colla co-cultured type I gel. Activation of lymphocytes was analyzed by secretion of γ -IFN and TNF- α (data not shown).

Apoptosis was analysed after 24h using the apoptosis profiler kit from R&D, detecting 40 related proteins. TRAIL-transduced lymphocytes in combination with bsAB EpcAMxCD3 showed an efficient activation of apoptosis in the 3D reconstructs (A). Detected proteins are shown in pixel density for PC-3 mixed with TRAIL-transduced lymphocytes +/- bsAb EpCAMxCD3 (B). BxPC3 gave similar results (data not shown).

TRAIL-transduced lymphocytes in combination with bsAB EpCAMxCD3 actively induce apoptosis in the

P., 100

y. The EpCAMxCD3 mce of EpCAM s did not affect s motility of lympho ever, duration of co ver, duration of contacts en lymphocytes and cells was three-times in the presence of MxCD3 (data not



tumor growth and morphology (C, D). CD45+ PBMCs even could be detected in the tumours after

23 days (B)

growth was strongly reduced in babb treated mice Compared to previous experiments, we observed a cyst formation in the BxPC-3 xenograft models in all groups with TRAIL-transduced lymphocytes (C).



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Results & Conclusion

taken. No metastasis could be found

We evaluated the anti-tumour efficacy of bispecific EpCAMxCD3 antibody linking tumour cells and T-lymphocytes. In NOD SCID mice, EpCAMxCD3 had a long serum half-life (t1/2 ~ 7 days). EpCAMxCD3 significantly reduced growth of BxPC-3 pancreatic and PC-3 prostate carcinoma xenografts. Since little apoptosis could be detected in these tumours on day 23 (TUNEL, Caspase 3), but lymphocytes (CD45+) could be detected – we assumed, that the time of observation for the mechanism resulting in the growth retardation and the cyst formation, was an earlier event. To further investigate the potential mechanisms of in vivo anti-tumour effects of EpCAMxCD3, we used a collagen gel 3D tumour reconstruct system, which closely resembled the tumour microenvironment. Therefore, to mimic the tumour situation, a mix of TRAIL transduced lymphocytes was co-cultured together with tumour cells and fibroblasts in collagen type I gel in vitro. In this setting apoptosis related proteins were expressed indicating an apoptosis induction in this system. To further prove the anti-tumour effect of the TRAIL-lymphocytes with the bsAb EpCAMxCD3, we performed kill assays incubating tumour cells with the bsAb loaded TRAIL-lymphocytes (data in preparation). With the previously demonstrated prolonged contacts between lymphocytes and tumour cells during the bsAb EpCAMxCD3, we could demonstrate a direct apoptosis effect through TRAIL-lymphocytes and bsAb in vitro to explain the observed anti-tumour effects in vivo. Overall, we provide a new combinatorial approach in which bsAb targeting EpCAM on the surface of tumour cells and genetically modified lymphocytes over expressing TRAIL could improve the therapy of advances prostate and pancreatic cancer in patients.